IVD

WizMag™ Total DNA

User Manual

Ver 2.0

REF W7000 | W7001 | W7002 | W7003

For *in vitro* diagnostic use

wizbiosolutions
THE INNOVATION OF GENOMICS

INTENDED USE

The WizMag™ Total DNA kit is designed to be used on the CLEO™ AP16 Nucleic Acid Extractor System for simple and easy purification of total DNA from various biological liquid samples including cultured cells, cell-containing media, body fluids and etc. Purified DNA is free of enzyme inhibitors and other contaminants and highly suited for downstream applications such as PCR-based or enzyme-based reactions.

KIT CONTENTS

Contents	7000	7001	7002	7003	Storage
No. of preparation	64	192	32	96	Room Temperature (15-25°C)
Prepacked 96-well plate	4 ea	12 ea	-	-	
Prepacked 6-well tube	-	-	32 ea	96 ea	
Plunger strip	8 ea	24 ea	8 ea	24 ea	
Buffer PKR	1.5 mL	6 mL	1 mL	6 mL	
Proteinase K *	14 mg x 2	42 mg x 2	14 mg	42 mg	
RNase A solution	340µL	1 mL	170 µL	500 µL	
Blank solution A	500 µL	500 µL	500 µL	500 µL	

This kit is delivered under ambient conditions. When being used immediately on arrival, all the components can be stored at room temperature (15 - 25 °C). But if the kit is going to be stocked for a long time, Proteinase K should be stored at 2 - 8°C for optimal conservation. Long exposure to heat sources can deteriorate the performance of the kit significantly.
* After dissolving, store proteinase K solution at 2 - 8°C for optimal conservations.

QUALITY CONTROL ANALYSIS

In accordance with Wizbiosolutions Inc. ISO 13485-certified Quality Management System, each lot WizMag™ Total DNA kit is tested against predetermined specifications to ensure consistent product quality.

RECONSTITUTION OF PROTEINASE K

Before the first experiment, dissolve completely Proteinase K with Buffer PKR, as indicated on the product label. Do not vortex when dissolving. Store the reconstituted Proteinase K solution at 2 - 8°C.

PRECAUTIONS

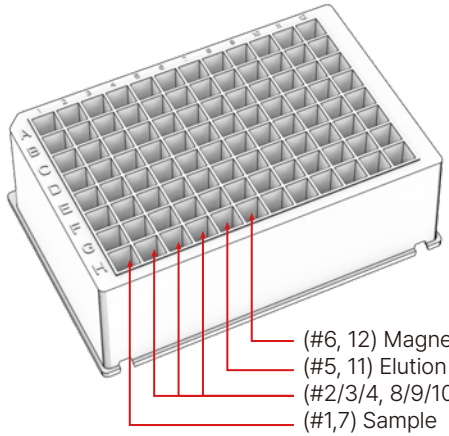
- This product is reserved exclusively for ***in vitro*** diagnostic purposes.
- Intended for single use only. Do not reuse.
- Check the expiration date on the box. Do not use it after the expiration date.
- Wear protective clothing, and use disposable gloves, goggles, and a mask.
- Do not eat, drink or smoke in areas where samples or test reagents are being used. Once you finish the test wash your hands.
- Specimens must be treated as potentially infectious as well as all reagents and materials that have been exposed to the samples and handled in the same

manner as an infectious agent.

- Regular decontamination of commonly used equipment is recommended, especially micropipettes and work surfaces.
- This product contains irritants that are harmful when in contact with skin or eyes, or inhaled or swallowed. Care should be taken when handling this product.
- Some of the reagents in the 96-well Plate contain chaotropic which can form highly reactive compounds when combined with bleach. Do NOT add bleach or acidic solutions directly to the sample-preparation waste.
- Any significant incidents related to the product should be notified to the competent authorities and manufacturers.
- Do not use it if the package is damaged.

COMPOSITION OF THE PRE-PACKED 96-WELL PLATE (W7000 | W7001)

A total of 16 samples can be simultaneously processed per plate.



Columns 7 - 12 in the right half of a 96-well plate have the same composition as columns 1 - 6 in the left half.

COMPOSITION OF THE PRE-PACKED 6-WELL STRIP (W7002 | W7003)

(#1) Sample
(#2/3/4) Washing
(#5) Elution
(#6) Magnetic bead

PROTOCOL

A. Setup of program (For CLEO™ AP16 & AP48 devices, preset program can be used.)

Edit and run the experiment program as follows:

No.	1	2	3	4	5	6	7	8
Well#	1	6	1	2	3	4	5	6
Step	Lysis	Beads	Bind	Wash	Wash	Wash	Elute	Discard
Wait time	-	-	-	-	-	-	03:00	-
Mix time	10:00	00:20	05:00	02:00	01:00	01:00	05:00	00:20
Mag time	00:00	00:25	00:30	00:25	00:25	00:25	00:45	-
Volume(µL)	700	200	700	750	750	750	100	200
Mixing speed	Fast	Med	Fast	Fast	Fast	Fast	Bottom	Medium
Collect speed	Normal	Strong	Strong	Strong	Strong	Strong	Strong	Normal
Temp	70°C					65°C		

B. Sample Preparation

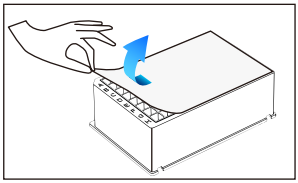
▶ Required optional material : 1x PBS

1. Cultured cells
   - Pellet up to 2×10^6 cells by centrifugation and resuspend the cell pellet in 200 µL of 1x PBS.

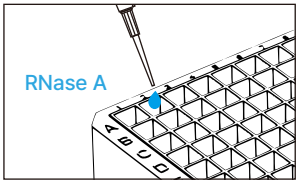
2. Cell-containing media or body fluids (Plasma, Serum, etc.)
   - Use up to 200 µL of specimen.
   - Adjust to 200 µL with 1x PBS if less.

3. Swab samples
   - Place the swab head in 400 µL of saline or PBS, and vortex vigorously for 30 seconds.
   - Use 200 µL of immersion solution.

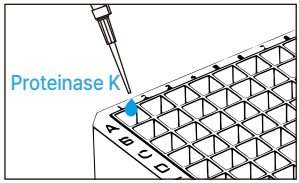
C-1. DNA extraction procedure (W7000, W7001)



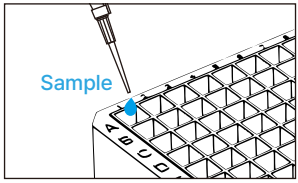
1. Carefully peel off the film of the 96-well Plate not to cross-contaminate.



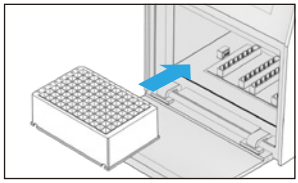
2. (Optional for RNA removal) Add 5 µL of RNase A solution into each second well (#2,8)
   - Alternatively, RNase A solution can be applied to the sample directly by 5 µL per sample.
   - It is recommended not to treat RNase A when the sample contains a few cells.



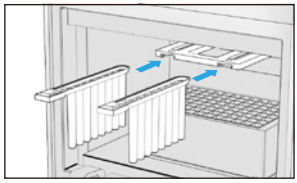
3. Add 20 µL of Proteinase K solution into the each first well (#1,7)



4. Add 200 µL of the sample into the each first well (#1,7)

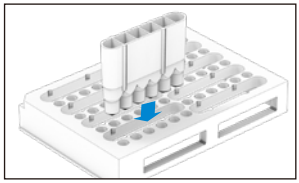


5. Mount the 96-well Plate on the CLEO™ AP16 carefully.

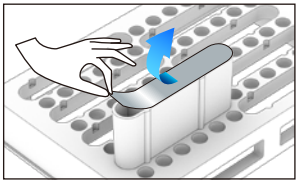


6. Insert a Plunger all the way into the socket above the 96-well Plate.

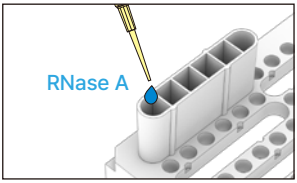
C-2. DNA extraction procedure (W7002, W7003)



1. Mount the 6-well Strip onto the Strip Adapter Plate.

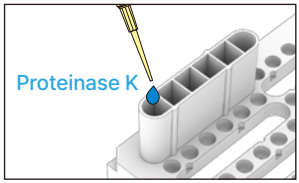


2. Carefully peel off the film of the 6-well Strip not to cross-contaminate.

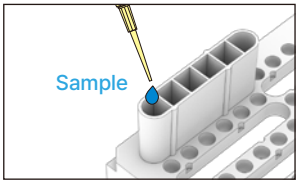


3-1.(Optional for RNA removal)
Add 5 µL of RNase A solution into each second well (#2,8)

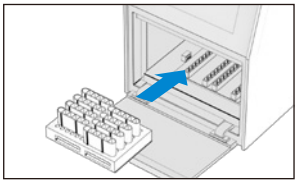
- Alternatively, RNase A solution can be applied to the sample directly by 5 µL per sample.
- It is recommended not to treat RNase A when the sample contains a few cells.



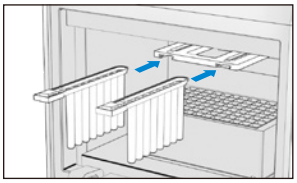
3-2. Add 20 µL of Proteinase K solution into the each first well (#1)



4. Add 200 µL of the sample into the each first well (#1).



5. Mount the 6-well Strip Adapter Plate on the CLEO™ AP16 carefully.



6. Insert a Plunger all the way into the socket above the Plate.

7. Close the front door of the instrument.
8. Select **MENU** ▶ **DNA** ▶ **Total DNA** on the screen.

8. Press **'RUN'** button on the screen.
9. After the alarm finishes, open the door and carefully remove the Plunger.
10. Detach the 96-well plate (or the Strip Adapter Plate) from the machine carefully.
11. Transfer the 60 - 80 µL eluate of each fifth well (#5, 11) into a new 1.5 mL centrifuge tube.

    NOTE : The volume of eluate can be decreased slightly during the process.
12. Dispose of 96-well Plates (or 6-well Strip) and Plunger used in the test according to local or national waste disposal methods.

TROUBLE SHOOTING GUIDE

Problem	Possible causes	Recommendations
Low or no recovery	Low cells in the starting sample	Some samples may have low population of cells. When the cell mass is very low, an addition of carrier RNA (5-10 µg/ sample) into the sample can be helpful for the recovery of DNA.
	Too many cells in the starting sample	The sample amount over the maximum capacity will lead to poor lysis of cells, resulting in significantly low recovery. Reduce the amount of starting sample.
Low purity	Too much sample amount used	Do not overload the sample. Keep the volume and the cell number of sample as procedure.
High A_{260}/A_{280} ratio	RNA contamination	RNA can inhibit some downstream enzymatic reactions, but not PCR itself. If RNA should be removed from the preparation, add RNase A solution into well#2 at DNA recovery procedure. Alternatively, RNase A solution can be applied to the sample directly by 5 µL per sample.
Degraded DNA	Starting sample is too old or improperly stored	Too old or improperly stored samples may have degraded DNA. Use a fresh sample.
Inconsisten recovery of DNA	Contamination between reagent wells	The reagent in the well may evaporate and form a deposit on the film during storage, which may cause contamination between wells when the film is removed. Prepacked plate or tube always should be stored at proper condition. Before removing the film of the plate or the tube, it is recommend-ed to shake off the deposit on the film with holding the plate or the tube tightly.

SYMBOL GLOSSARY

REF	Catalogue number		Manufacturer		Use-by date
LOT	Batch code		Do not re-use		Temperature limitation
IVD	in-vitro diagnostic use		Instructions for use		Keep away from sunlight
	Contents sufficient for <n> tests		Caution		Keep dry
	Do not use if package is damaged	UDI	Unique Device Identification		

ORDERING INFORMATION

Product	Cat No.	Package	Note
WizMag™ Total DNA	W7000	64 Prep	16 prep/run
	W7001	192 Prep	
	W7002	32 Prep	Single prep
	W7003	96 Prep	
CLEO™ AP16 Nucleic acid Extractor	CL2016	1 system	1-16 sample
CLEO™ AP48 Nucleic acid Extractor	CL2048	1 system	1-48 sample

MFDS License No. : IVD-23-834

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THE INNOVATION OF GENOMICS

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